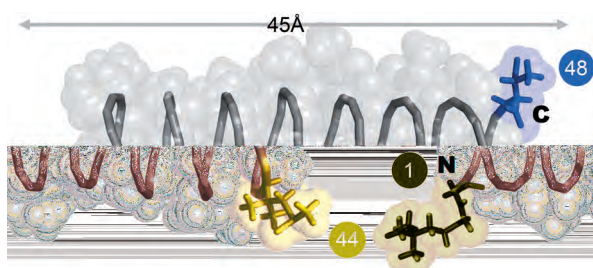


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# Selective Modification of the N-Terminal Structure of Polytheonamide B Significantly Changes its Cytotoxicity and Activity as an Ion Channel

Naoki Shinohara, Hiroaki Itoh, Shigeru Matsuoka, and Masayuki Inoue<sup>\*[a]</sup>

Polytheonamide B (**1**; Scheme 1a) is an extremely cytotoxic natural product ( $IC_{50} = 0.098$  nM, mouse P388 leukemia cells), and it is the largest non-ribosomal peptide described to date ( $MW = 5030$  Da).<sup>[1,2]</sup> The linear-chain polypeptide **1** is formed of an N-terminal 5,5-dimethyl-2-oxohexanoate group (Ncap) and 48 amino acid residues of alternating D- and L-chirality, and the secondary structure was recently reported to be a remarkable  $\beta^{6.3}$ -helix with a length of approximately 45 Å (Figure 1).<sup>[3]</sup>



**Figure 1.** The  $\beta^{6.3}$ -helix structure of polytheonamide B (**1**) determined using NMR (PDB: 2RQO<sup>[3]</sup>). Labels N and C denote the N and C termini, respectively. Stick models of the Ncap and residues 1, 44, and 48 are shown. Colors correspond to those in Scheme 1.

This helical tube structure is believed to function as a transmembrane ion channel in biological settings, as the hydrophilic pore, which is 4 Å in diameter, creates a path for ion flow.<sup>[4]</sup> Planar bilayer experiments demonstrated that monomeric **1** forms a monovalent-cation-selective channel.<sup>[5]</sup> Therefore, cytotoxic peptide **1** not only poses a synthetic challenge, but also provide a unique structural platform for designing cytotoxic molecules and synthetic transmembrane channels.<sup>[6]</sup>

We recently reported the total synthesis of polytheonamide B,<sup>[7]</sup> as well as the synthesis of nine substructures of 7 to 37 amino acid residues. We found that all of the substructures are at least 10000-times less cytotoxic than the parent compound.<sup>[8,9]</sup> These studies suggest that the overall membrane spanning property of **1** is important for its biological activity. To further characterize the structure–function relationship of **1**, we planned to site-specifically modify monomer units of synthetic polytheonamide B and evaluate the biological activity of the newly generated analogues. We hypothesized that this type of molecular editing<sup>[10]</sup> would enable us to decipher the structural elements that are responsible for the unique biological

functions of **1**. In the first set of experiments, the 44th, N- and C-terminal residues were selected as the modification points. Here, we report the synthesis of seven derivatives of polytheonamide B and discuss the significant effect of N-terminal modification on their cytotoxicity and ability to act as ion channels in vitro.

Achieving selective and precise alteration of a complex structure such as that of polytheonamide B necessitates the use of highly chemoselective reactions, since the numerous native functional groups of **1** can cause undesired reactions. For instance, attempted methyl ester formation (**1**→**2**) at the C terminus resulted in generation of multiple methylated products when **1** was exposed to trimethylsilyldiazomethane (TMSCHN<sub>2</sub>). Thus, methyl ester **2** was alternatively produced by treating **1** with 1-hydroxybenzotriazole (HOBt) and *N,N*-diisopropylcarbodiimide (DIPCDI) in *N,N*-dimethylformamide (DMF) and methanol (Scheme 1b). The sulfoxide moiety of the 44th amino acid residue of **1** was chemoselectively oxidized to the sulfone group of **3** in 69% yield.<sup>[11,12]</sup> Then, a series of the N-terminal-modified analogues **5**–**8** was prepared from amine **4**, which was produced by taking advantage of the 1,2-dicarbonyl structure of Ncap in peptide **1**. Specifically, reaction between **1** and benzene-1,2-diamine under Dixon conditions<sup>[13]</sup> led to the formation of **4** and **9** without cleavage of the other numerous amide bonds. Amine **4** in turn was transformed into acetamide **5**, octanamide **6**, and palmitamide **7**, as well as trimethyl ammonium derivative **8** by applying the corresponding anhydride or mixed anhydrides as shown in Scheme 1b.<sup>[14]</sup>

The cytotoxicity of synthetic derivatives **2**–**8** was assessed using a colorimetric assay and 2,3-bis-(2-methoxy-4-nitro-5-sulphophenyl)-2H-tetrazolium-5-carboxanilide (XTT) in mouse P388 leukemia cells (Table 1).<sup>[15]</sup> While the cytotoxicity of methyl ester analogue **2** was comparable to the parent **1**, sulfone **3** was fivefold less toxic than **1**. Remarkably, the results indicate that the degree of toxicity is dependent upon the oxidation state of a single sulfur atom, suggesting that the highly polar

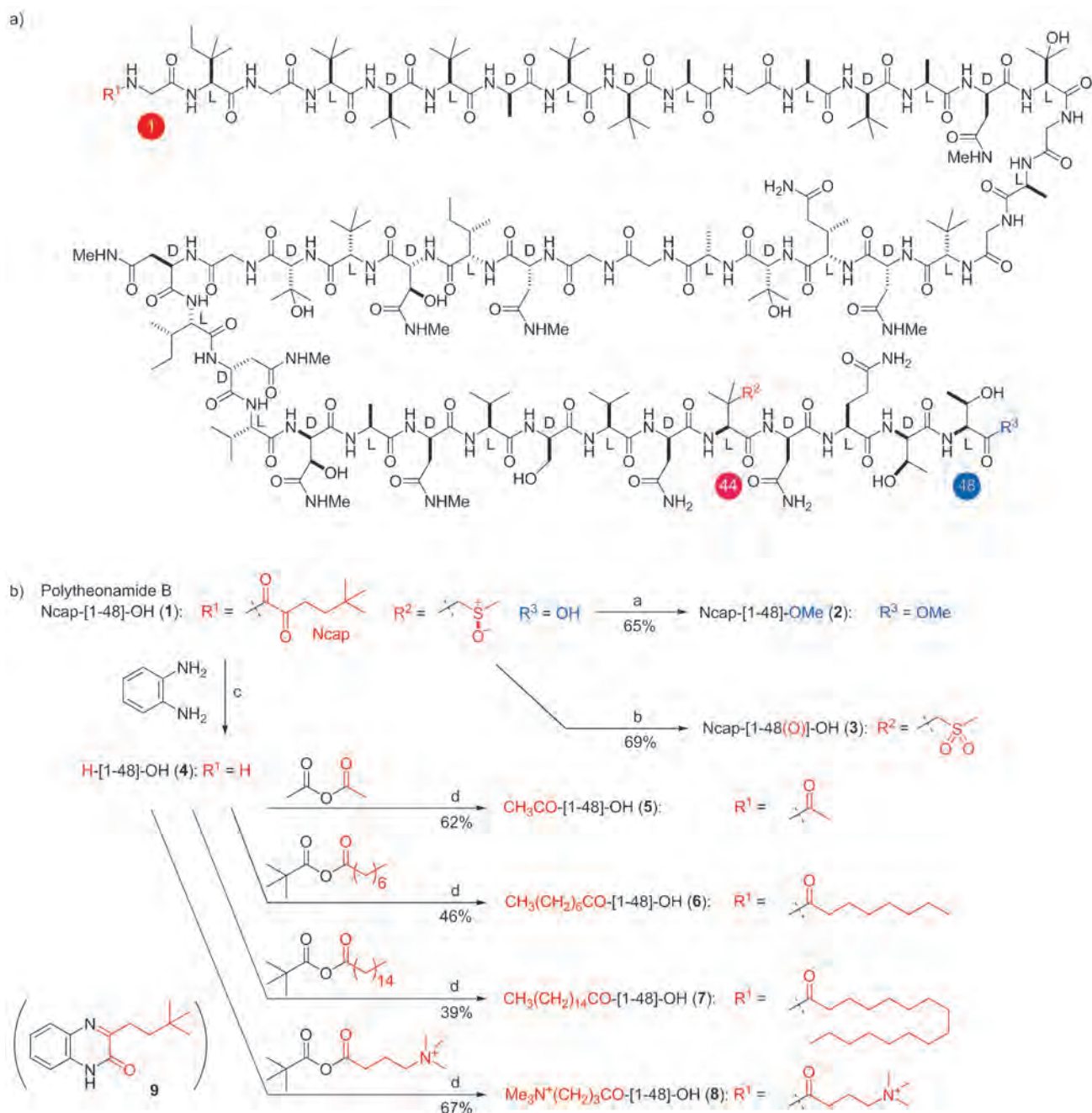
**Table 1.** Cytotoxicity of polytheonamide B (**1**) and analogues **2**–**8**.

Synthetic peptides	$IC_{50}$ <sup>[a]</sup> [nM]
Ncap-[1-48]-OH ( <b>1</b> )	0.098
Ncap-[1-48]-OMe ( <b>2</b> )	0.12
Ncap-[1-48(O)]-OH ( <b>3</b> )	0.46
H-[1-48]-OH ( <b>4</b> )	24
CH <sub>3</sub> CO-[1-48]-OH ( <b>5</b> )	47
CH <sub>3</sub> (CH <sub>2</sub> ) <sub>6</sub> CO-[1-48]-OH ( <b>6</b> )	0.50
CH <sub>3</sub> (CH <sub>2</sub> ) <sub>14</sub> CO-[1-48]-OH ( <b>7</b> )	0.038
Me <sub>3</sub> N <sup>+</sup> (CH <sub>2</sub> ) <sub>3</sub> CO-[1-48]-OH ( <b>8</b> )	250

[a]  $IC_{50}$  values were determined from the results of growth inhibition assays (XTT method) performed on mouse P388 leukemia cells.

[a] Dr. N. Shinohara, H. Itoh, Dr. S. Matsuoka, Prof. Dr. M. Inoue  
Graduate School of Pharmaceutical Sciences, The University of Tokyo  
7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033 (Japan)  
E-mail: inoue@mol.f.u-tokyo.ac.jp

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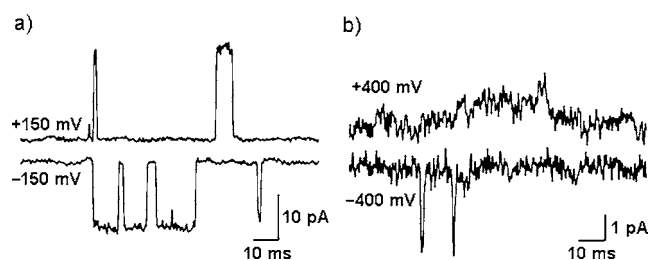
**Scheme 1.** a) Structure of polytheonamide B (1). b) The synthesis of polytheonamide analogues 2–8. *Reagents and conditions:* a) HOBT, DIPCPI, DMF/MeOH (1:1), 0→27 °C; b) oxone, *n*PrOH/H<sub>2</sub>O (3:1), 30 °C; c) 2 M NaOAc/HOAc (pH 4), DMF, 40 °C; d) pyridine (for 5 only), DMF, 27 °C. Abbreviations: *N,N'*-diisopropylcarbodiimide (DIPCPI), *N,N*-dimethylformamide (DMF), 1-hydroxybenzotriazole (HOBT), 5,5-dimethyl-2-oxohexanoic acid (NcapOH), 2KHSO<sub>5</sub>·KHSO<sub>4</sub>·K<sub>2</sub>SO<sub>4</sub> (oxone).

nature of the sulfoxide group of the 44th residue plays a favorable role in the toxicity.<sup>[12,16]</sup>

Structural modification of the N terminus had an even more profound effect on cytotoxicity. The less hydrophobic compounds amine 4 and acetamide 5, and the comparably hydrophobic octanamide 6, were 240-fold, 480-fold, and fivefold less toxic than the parent 5,5-dimethyl-2-oxohexanamide 1, respectively, whereas the more hydrophobic palmitamide 7 was even more cytotoxic than 1. The apparent correlation between hydrophobicity and cytotoxicity among compounds 1, 4, 5, 6

and 7 indicates that the size of the hydrophobic N-terminal moieties is critical for their cytotoxicity. The most drastic decrease in cytotoxicity (2500-fold) was exhibited when the Ncap group of 1 was switched to a trimethyl ammonium structure (compound 8). The ammonium cation appears to have a strongly negative impact on the intrinsic potency of 1. These results suggest that hydrophobic substitution at the N terminus is required for targeting cell membrane as well as orienting 1 for effective membrane insertion, where it exerts its cytotoxic activity.

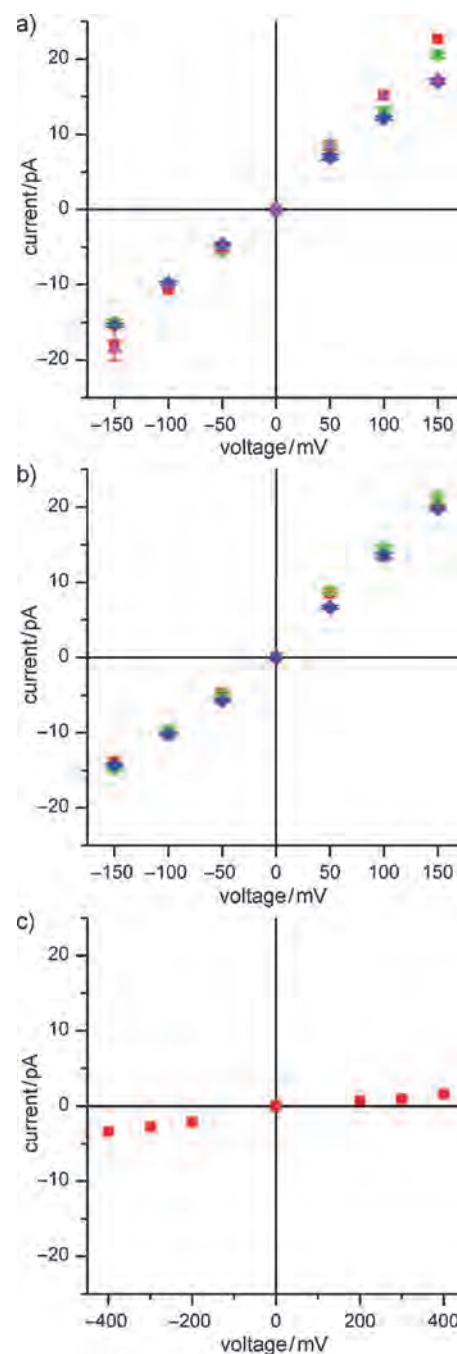
To compare the function of the ion channel formed by polytheonamide B derivatives **2–8** with that of **1**, transmembrane proton (single-channel) currents were recorded by patch clamp assay in acidic aqueous solution using an artificial planar lipid bilayer.<sup>[17–19]</sup> The results showed that channels with distinct open and closed states formed immediately in the planar lipid bilayer after addition of **1** and derivatives **2–7** (Figure 2a). In contrast, virtually no channel current was observed when trimethyl ammonium derivative **8** was added to the planar lipid bilayer (Figure 2b).



**Figure 2.** Single-channel current recordings for analogues **7** and **8** made with the Port-a-Patch patch clamp system from Nanion Technologies. Traces show proton current flow through a channel formed by a) **7** or b) **8** in a planar diphytanoyl phosphatidylcholine (DPhPC) bilayer in 0.1 M HCl.

Accordingly, the current–voltage curves for all polypeptide analogues except **8** were comparable, with a constant conductance in the range of  $\pm 150$  mV (Figure 3a,b). Analogue **8** blocked the proton current in a higher voltage range ( $\pm 400$  mV; Figure 3c). Our data indicate that the structural differences in analogues **2–7** are inconsequential as far as impacting the *in vitro* activity of the proton channel formed by **1**. In contrast, the mere presence of the ammonium group in **8** almost completely eliminates the proton channel activity. It is possible that the positively charged group at the N terminus of **8** mimics a monovalent cation and thus strongly binds to the cation entrance of the tubular structure (see Figure 1), resulting in a “closed end” and so blockade of ion passage.<sup>[20]</sup> The negligible ion channel activity of **8** correlates with its significantly lower cytotoxicity of **8** as compared with compounds **1–7**, suggesting that the two activities are related. Overall, we have demonstrated that both the cytotoxicity and ion channel activity can be dramatically altered by varying the structure of the N terminus.

In conclusion, polytheonamide B (**1**) was structurally modified at the 44th, N- or C-terminal amino acid residue to generate derivatives **2–8**, which were evaluated in cytotoxicity assays and single-channel recording experiments. We have demonstrated for the first time that the hydrophobicity of the N-terminal structures of **3–7** correlates strongly with the cytotoxicity exhibited by the compound, and that palmitamide **7** is more potent than the parent natural product (**1**) despite its similar behavior as a proton channel. Furthermore, the drastic deactivating effect of the trimethyl ammonium moiety of **8** on both the cytotoxicity and proton current was revealed. These data suggest that hydrophobic N-terminal moieties function as membrane-targeting anchors critical for cytotoxicity and that



**Figure 3.** Current (*I*)–voltage (*V*) curves for proton channels formed by a) Ncap-[1–48]-OH (**1**: ■), Ncap-[1–48]-OMe (**2**: ●), Ncap-[1–48(O)]-OH (**3**: ◆), H-[1–48]-OH (**4**: ▲), b) MeCO-[1–48]-OH (**5**: ■), Me(CH<sub>2</sub>)<sub>6</sub>CO-[1–48]-OH (**6**: ●), Me(CH<sub>2</sub>)<sub>14</sub>CO-[1–48]-OH (**7**: ◆), and c) Me<sub>3</sub>N<sup>+</sup>(CH<sub>2</sub>)<sub>3</sub>CO-[1–48]-OH (**8**: ■).

an ammonium cation in the same position acts as an intramolecular channel blocker. The results highlight the functional roles of the 44th, N-, and C-terminal residues on the cytotoxicity and channel activities of these polypeptides, and should facilitate efforts to rationally design tailor-made transmembrane ion channels based on the structure of polytheonamide B.

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**Keywords:** cytotoxic agents · natural products · nonribosomal peptides · structure–activity relationships · synthetic ion channels

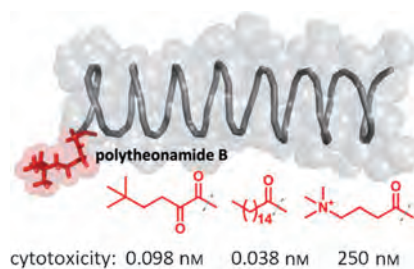
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## COMMUNICATIONS

**Chemical point mutation:** Polytheonamide B is a naturally occurring polypeptide containing 48 amino acids. It both displays potent cytotoxicity and acts as a monovalent cation channel in vitro. Chemoselective methods to modify the 44th, N-, and C-terminal residues of the natural product have been developed, and evaluation of the resultant derivatives suggests that the intrinsic activities of the peptide can only be altered by switching its N-terminal substitution.



*N. Shinohara, H. Itoh, S. Matsuoka, M. Inoue\**

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**Selective Modification of the N-Terminal Structure of Polytheonamide B Significantly Changes its Cytotoxicity and Activity as an Ion Channel**

